

Inflammation meets cancer, with NF- κ B as the matchmaker

Yinon Ben-Neriah¹ & Michael Karin²

Inflammation is a fundamental protective response that sometimes goes awry and becomes a major cofactor in the pathogenesis of many chronic human diseases, including cancer. Here we review the evolutionary relationship and opposing functions of the transcription factor NF- κ B in inflammation and cancer. Although it seems to fulfill a distinctly tumor-promoting role in many types of cancer, NF- κ B has a confounding role in certain tumors. Understanding the activity and function of NF- κ B in the context of tumorigenesis is critical for its successful taming, an important challenge for modern cancer biology.

In 1863, Rudolf Virchow, the founder of modern pathology, noted leukocytes in neoplastic tissues and made a connection between inflammation and cancer. He suggested that the “lymphoreticular infiltrate” reflected the origin of cancer at sites of chronic irritation. In the dawn of the 20th century, Katsusaburo Yamagiwa showed that repeated painting of coal tar onto rabbits’ ears causes carcinomas. Later, in the 1940s, using repeated application of tar or croton oil onto the skin, Peyton Rous and Isaac Berenblum introduced the concept of tumor promotion, a pathogenic process distinct from tumor initiation. The early studies of Yamagiwa on the pathogenesis of gastric carcinoma led to his belief that chronic gastric ulcers have a major role in the development of stomach cancer. In 1911, he established principles that later led him to uphold the irritation theory of cancer. Seventy years later, Barry Marshall and Robin Warren proved that gastritis is caused by infection with *Helicobacter pylori*, now thought to be a major cause of many stomach cancers. Although those classical studies pointed to an association between inflammation and cancer, the mechanistic basis of this relationship emerged subsequently, with the transcription factor NF- κ B serving as the major lynchpin. Here we review the function of NF- κ B in linking inflammation to cancer. However, rather than providing a detailed summary of the inflammation-cancer connection, this review is focused on certain outstanding issues, such as the relationship between NF- κ B activation and abnormal growth signaling, the interaction between the positive and negative roles of NF- κ B in the control of inflammatory responses and how these opposing functions affect tumor development and progression. More extensive reviews of the inflammation-cancer field have been published elsewhere^{1–5}.

Evolutionary linkage of NF- κ B and abnormal growth

Inflammation is a manifestation of innate immunity, a fundamental protective response that is conserved in all multicellular animals⁶. The emergence of multicellular life forms required new means for defending these slow-growing organisms from rapidly growing invading pathogens and for preventing the fusion of genetically distinct conspecific organisms⁷. Shortly after the discovery of NF- κ B, it was postulated that it ‘plays the first violin’, if it is not the ‘conductor’ of inflammatory responses^{8,9}. Although inflammation can be induced in the absence of NF- κ B¹⁰, that is rarely a physiological occurrence, which possibly reflects the need for the transcription factor not only for amplification and maintenance of inflammation but also for ‘tuning down’ and curtailing inflammation, to preserve tissue function once the inflammation is no longer needed¹¹.

The innate immune system is well suited for detecting pathogens and foreign bodies and reacts to them by producing and releasing immune effectors and activated cells that either contain or eliminate the pathogen. An intricate signaling system composed of sensors, signal-processing and signal-transducing elements, and myriad effector molecules, from reactive oxygen species (ROS) and antibacterial peptides to diffusible regulators of immunity (cytokines and chemokines) was constructed for that purpose through evolution. The basic innate immunity scheme is remarkably well conserved both structurally and functionally. Thus, the main classes of pathogen sensors, Nod-like receptors and Toll-like receptors (TLRs) and interleukin 1 (IL-1) receptors, as well as the signal transducers and amplifiers IRAK, MyD88, TRAF and IKK and the transcriptional regulator Rel (NF- κ B), are present even in the most primitive metazoans, sponges, sea anemones, hydra and jelly fish^{12–15}. Notably, the RIG-I-like receptor family, another major arm of innate immunity that controls viral infection through the interferon response, originated much later than Nod-like receptors and TLRs, possibly only in vertebrates¹⁶.

Given its considerable conservation noted above, innate immunity can be considered a hallmark of multicellularity, one of the following six essential principles of metazoan life: regulated cell replication and growth; programmed cell death; cell-cell and cell-matrix adhesion; regulated developmental processes; cell type specialization; and alleloreactivity and

¹Lautenberg Center for Immunology, Institute for Medical Research-Israel-Canada, Hebrew University-Hadassah Medical School, Jerusalem, Israel.

²Laboratory of Gene Regulation and Signal Transduction, Department of Pharmacology and Cancer Center, School of Medicine, University of California, San Diego, La Jolla, California, USA. Correspondence should be addressed to Y.B.-N. (yinion@cc.huji.ac.il) or M.K. (karinoffice@ucsd.edu).

innate immunity¹⁷. Although NF- κ B and inflammation are commonly associated with the last principle, in fact all the principles noted above are affected by NF- κ B: it contributes to induction of proliferative genes^{18,19}; it regulates genes encoding antiapoptotic molecules²⁰; it controls the expression of diverse adhesion molecules^{21,22}; it drives and supports developmental processes from lymphocyte differentiation to mammary gland development^{23,24}; and it even has a role in cell specialization, as in driving Schwann cells to myelinate²⁵.

As many of those diverse functions go awry in tumorigenesis, it is interesting to trace the evolutionary origin of the inflammation-cancer link. Two remarkable examples are tumor promotion in *Drosophila* larva, in which hemocyte tumor necrosis factor (TNF) enhances tumor growth and stimulates the invasive migration of cells with mutation of the genes encoding the oncoprotein Ras and the tumor suppressor Scribble²⁶; likewise, leukocyte-trophic effects are needed for the promotion of melanoma growth in zebrafish larva²⁷. Additional hints of a relationship between the innate response and abnormal cell growth can be found even earlier in metazoan evolution, perhaps as early as corals, which frequently develop abnormal growths resembling tumors²⁸. In most coral specimens examined, these malformations are directly attributable to effects of predation or other physical injury. These malformations have many features in common with neoplasms, including failure of natural growth control and breakdown of the normal symmetrical pattern, and overall they resemble adenomatous polyps of the human colon^{29,30}. Deep-water corals are repeatedly preyed on by fish and other carnivores, which injure the soft parts of the coral²⁸. This results in microbial and viral infections that trigger an inflammatory reaction that promotes regenerative proliferation and abnormal growth. Harold Dvorak described tumors as “wounds that won’t heal,” pointing to many similarities between the activity of a cancerous tumor growth and the process of wound healing³¹. We thus speculate that NF- κ B-orchestrated innate immunity has been entwined with growth control from the early days of multicellularity. If infection is effectively controlled, then the inflammatory response is promptly resolved with no perturbation of tissue growth. Repeated infection, however, may result in tissue loss and a protracted inflammatory response with attempt to restore the lost tissue and, thus, as in damaged corals, may end in abnormal growth.

There is mounting evidence today that many tumors are propagated by means of cancer stem cells, rare cells in tumors with indefinite capacity for self-renewal³². Other tumors might arise from normal tissue stem cells or from tissue stem cells that were transformed to become cancer stem cells^{33,34}. If that holds true, tissue stem cells should be closely guarded against infectious and chemical genotoxic insults and, at the same time, might be particularly vulnerable to deregulated innate immunity. A notable example is the close proximity of intestinal stem cells to Paneth (CD24⁺) cells in the small bowel and similar CD24⁺ cells in the colon³⁵, the innate immune guardians of the gut epithelium. Indeed, evidence suggests that heterotypic Paneth cell-stem cell interactions have an important role in controlling stem-cell renewal³⁵. At the far end of this innate immunity-growth control equation, abnormal growth and cancer may be found, as in the transformation of intestinal crypt stem cells into microadenomas³⁴. Moreover, a remarkable synergy has been observed between bacterial infection and oncogenic mutations in *Drosophila* gut that drives abnormal enterocyte growth and dysplasia. Infection of *Drosophila* gut by *Pseudomonas aeruginosa*, a human opportunistic pathogen, induces intestinal damage, apoptosis and compensatory proliferation, which on a background of mutated *Ras* is excessive, with polarity loss, resembling a tumor³⁶. We hypothesize that microflora-induced innate immune responses in intestinal stem cells or their niche

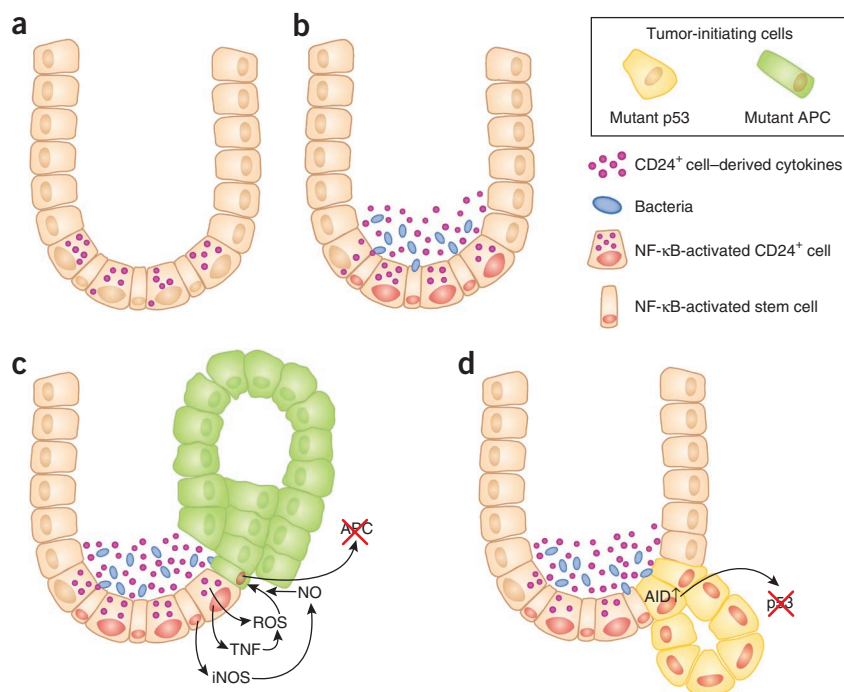
(composed mainly of CD24⁺ cells³⁵), possibly in conjunction with epigenetic alterations³⁷, may drive an abnormal proliferative response, which after further mutagenesis may generate tumor-initiating stem cells (Fig. 1).

Another striking example of the equivalence of excessive innate immunity activation and abnormal growth signaling is the discovery that deletion of *NFKBIA*, which encodes the NF- κ B inhibitor I κ B α , is a rather frequent oncogenic event in glioblastoma tumors³⁸. The occurrence of this mutation is mutually exclusive with the common glioblastoma amplification of epidermal growth factor receptor (EGFR), which indicates that activation of NF- κ B can replace aberrant EGF signaling as an oncogenic factor. Likewise, lung cancer cells with mutant EGFR are particularly sensitive to inhibition of NF- κ B, and NF- κ B activation through deletion of I κ B α rescues EGFR-mutant lung cancer cells from the cytotoxic effects of the EGFR kinase inhibitor erlotinib³⁹. Hence, whether innate immunity or a trophic control factor is deranged, the outcome could be similar; that is, abnormal growth.

Pro- and anti-inflammatory functions of NF- κ B

Since the realization that NF- κ B is an inducible rather than cell type-specific transcription factor that responds to proinflammatory cytokines and microbial products, NF- κ B has been thought of as the key regulator of inflammation⁸. Indeed, NF- κ B-binding sites have been found in the promoters of most genes encoding cytokines and chemokines⁴⁰, and NF- κ B activation has been shown to be essential for their induction in response to immune and inflammatory challenges⁴¹. Although originally NF- κ B was associated exclusively with immune and inflammatory cell function, the realization that such transcription factors also have essential roles in epithelial tissues, as in coordinating antimicrobial immunity and maintaining barrier function in the gastrointestinal system, soon followed^{42,43}. Furthermore, activated or nuclear NF- κ B proteins have been detected in many chronic inflammatory conditions, including inflammatory bowel disease^{44,45}, rheumatoid arthritis⁴⁶ and psoriasis⁴⁷. These diseases respond to anti-TNF therapy⁴⁸, and the role of NF- κ B in activating *TNF* transcription has been established¹⁰. Correspondingly, mouse models of inflammatory bowel disease^{45,49}, rheumatoid arthritis^{9,50–52} and other inflammatory diseases respond positively to inhibitors of NF- κ B, which has raised enthusiasm about NF- κ B and IKK β as therapeutic targets in chronic inflammation and autoimmunity⁵³. Even under acute inflammatory conditions, NF- κ B is expected to have an important causal role, as genetic polymorphisms that potentiate NF- κ B activation increase mortality due to sepsis⁵⁴. With that in mind, it was a big surprise and a disappointment when inhibition of NF- κ B was found to increase or even cause inflammation under some circumstances. One of the earliest alarming observations was greater susceptibility to chemical-induced colitis in mice lacking IKK β in intestinal epithelial cells (IECs)⁵⁵. An even more severe and spontaneous inflammatory condition has been observed in mice devoid of IKK γ (NEMO) in IECs; these mice have an almost complete loss of NF- κ B activity in these cells⁵⁶. Likewise, ablation of IKK γ (NEMO) in mouse keratinocytes results in the development of a psoriasis-like inflammatory condition, which, surprisingly, is dependent on TNF⁵⁷. Initially, those findings were attributed mainly to the absence of NF- κ B-mediated cell-survival functions at epithelial surfaces, which serve as barriers that prevent the exposure of underlying tissue macrophages and dendritic cells to commensal bacteria. In support of that interpretation, a variety of genes encoding molecules involved in the maintenance of epithelial layer integrity, in addition to genes encoding standard antiapoptotic molecules, have been found to be under the control of NF- κ B⁵⁷. However, the real ‘clincher’ was provided by studies

Figure 1 Hypothetical model for the generation of colorectal tumors as a result of interplay among intestinal crypt microflora NF- κ B activation, and mutagenesis mechanisms in intestinal stem cell. Encounters of bacteria with stem cells and their niche (composed mainly of Paneth-like CD24⁺ cells³⁵ (granule-filled cells)) at the bottom of the colonic crypts may induce activation of NF- κ B in Paneth cells and stem cells. NF- κ B activation results in the release of cytokines and the production of ROS and nitric oxide (NO), as well as the upregulation of activation-induced cytidine deaminase (AID) in the stem cells¹⁴⁹, which all results in stem cell mutagenesis. Further activation of NF- κ B in tumor-initiating cells supports their survival. (a) A normal colonic crypt with CD24⁺ cells and stem cells (thin columnar cells) at the bottom. (b) Bacteria-loaded crypt, which results in NF- κ B activation in CD24⁺ cells and stem cells (red nuclei) and the release of cytokines and enzymes. (c) NF- κ B-mediated production of ROS and nitric oxide, which results in mutagenesis of the gene encoding adenomatous polyposis coli (APC) in an intestinal stem cell and adenoma growth³⁴. iNOS, inducible nitric oxide synthase. (d) NF- κ B-induced upregulation of activation-induced cytidine deaminase (AID), which results in mutagenesis of the gene encoding p53, dysplasia and invasion¹⁵⁰, typical of colorectal cancer associated with inflammatory bowel disease¹⁵¹.



of mice with inducible deletion of the gene encoding IKK β (IKK β^{Δ} mice) in cells responsive to type I interferon, which include myeloid progenitors, mature myeloid cells, lymphocytes, fibroblasts and epithelial cells in tissues in which large amounts of type I interferon are produced. IKK β^{Δ} mice are hypersusceptible to septic shock induced by either lipopolysaccharide or bacterial infection⁵⁸. Similar results have been obtained by repetitive treatment of normal mice with a specific IKK β inhibitor⁵⁸. Even without any challenge, mice treated with an IKK β inhibitor or IKK β^{Δ} mice develop progressive and devastating neutrophilia due to more production of IL-1 β by NF- κ B-deficient monocytes and macrophages⁵⁹. Such experiments have shown that in addition to its proinflammatory function, NF- κ B has a direct anti-inflammatory effect; that is, inhibition of inflammasome-dependent caspase-1 activation⁵⁸ (Fig. 2). Although the mechanism of inflammasome inhibition by NF- κ B is not entirely clear, it is probably related to NF- κ B-induced expression of antiapoptotic proteins, such as PAI-2 and Bcl-x_L (refs. 58,59). The IL-1 β released by NF- κ B-deficient macrophages and monocytes enhances the proliferation of granulocytic progenitors and increases the survival of mature neutrophils⁶⁰. Although the resulting neutrophilia compensates for the loss of NF- κ B and allows IKK β^{Δ} mice to resist certain microbial infections as well as (or even better than) their wild-type counterparts, it eventually results in the inflammatory destruction of tissues, which can be prevented by inhibition of IL-1 β signaling⁶⁰. Although such results seem to suggest that a more effective inhibition of inflammation can be achieved by combining inhibitors of IKK β and IL-1 β , it should be noted that IKK β^{Δ} mice that are also deficient in the IL-1 receptor show a complete lack of innate immunity⁶⁰ and that the combined use of anti-TNF and anti-IL-1 drugs in humans results in much greater risk of infection⁶¹. Notably, IL-1 β , whose production is subjected to both positive and negative controls by NF- κ B, may function as a potent tumor promoter in some types of cancer⁶².

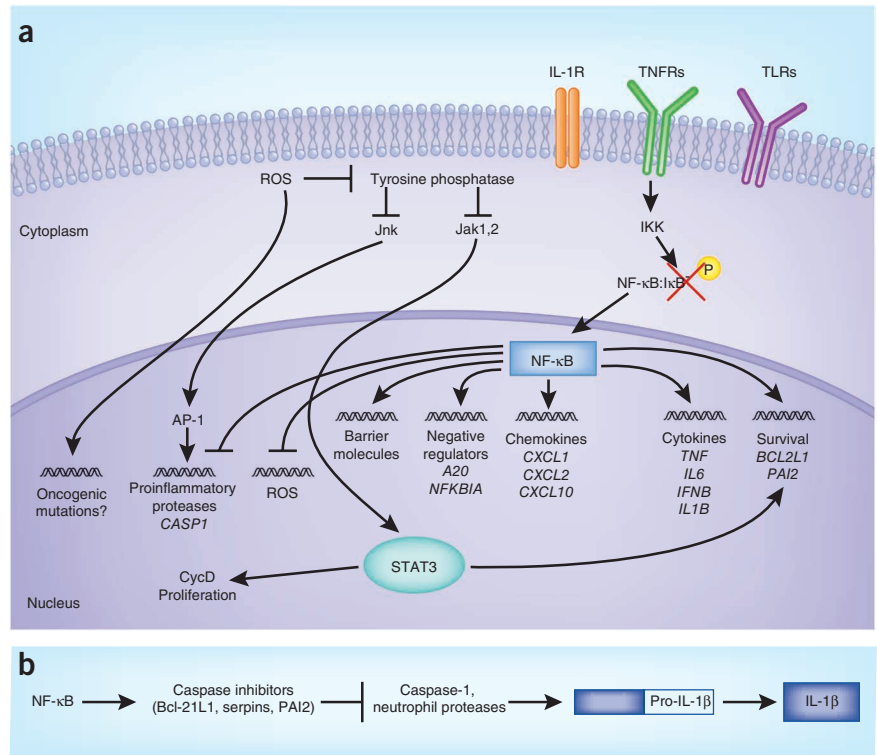
The inflammatory response is a complex physiological host-defense system. In addition to being important for clearing foreign intruders,

inflammation is important for the turnover and repair of damaged tissues. To function properly, the inflammatory response must be self-limiting and self-resolving. NF- κ B orchestrates both the initiation of inflammation and its resolution^{11,63}. In addition, part of the self-limiting nature of the inflammatory response is due to the existence of NF- κ B-dependent feedback loops, such as those that entail the induction of I κ B α and the ubiquitin-editing enzyme A20 (ref. 64). Thus, although inhibition of NF- κ B often attenuates inflammation, under somewhat different circumstance or at a different site it can aggravate or even cause inflammation. The latter outcome often becomes particularly prominent under conditions of tissue injury^{65,66}.

Pro- and anti-tumorigenic roles of NF- κ B in malignant cells

A potential role for NF- κ B in oncogenesis was already evident in the discovery of the retroviral oncogene *v-Rel* as the homolog of the gene encoding c-Rel, one of the NF- κ B subunits⁶⁷. Subsequently, mutations in genes encoding NF- κ B subunits or I κ B proteins, most prominent among which were chromosomal translocations in *NFKB2*, were identified in a variety of hematological malignancies⁶⁷⁻⁷⁰. However, the number of tumors with activated nuclear NF- κ B is much larger than the subfraction of malignancies with confirmed mutations in NF- κ B- or I κ B-encoding genes. Such observations led to the proposal that some of the NF- κ B activation seen in cancer is due to mutations that affect components of signaling pathways that activate NF- κ B or is the result of exposure to inflammatory cytokines in the tumor microenvironment⁷¹. Indeed, upstream mutations that cause NF- κ B activation were first detected in MALT lymphomas, a group of tumors that arise through chronic antigenic stimulation of mucosal-associated lymphoid tissue (MALT). Common MALT lymphoma mutations include chromosomal translocations that increase expression of the adaptors Bcl-10 (ref. 72) and MALT1 (ref. 73) and lead to constitutive assembly of the Carma-1-Bcl-10-MALT1 complex, whose normal function is activation of IKK-NF- κ B, downstream of antigen receptors^{74,75}. Constitutive activation of NF- κ B results in greater proliferation

Figure 2 Pro- and anti-inflammatory functions of NF- κ B and their relationship to tumorigenesis. (a) Activation of NF- κ B downstream of TNF receptors (TNFRs), TLRs and the IL-1 receptor (IL-1R) results in the induction of genes encoding prosurvival and pro-proliferative molecules, cytokines and chemokines. The products of such genes contribute to inflammation and tumor development. However, NF- κ B activation also promotes tissue integrity through the induction of genes encoding barrier molecules, protease inhibitors and antioxidants. Such molecules can suppress tumor development. By inducing the expression of antioxidant proteins, NF- κ B also prevents the accumulation of pro-tumorigenic ROS and can induce DNA damage and genomic instability and lead to the activation of pro-tumorigenic transcription factors, such as STAT3 and AP-1. (b) A particularly intriguing NF- κ B target gene encodes pro-IL-1 β , which is processed by caspase-1 or neutrophil protease to the key proinflammatory and tumor-promoting cytokine IL-1 β . Notable, while promoting pro-IL-1 β expression, NF- κ B negatively controls its processing to mature IL-1 β through the induction of various protease inhibitors.



and survival of B lymphocytes, which leads to their uncontrolled accumulation even after the initiating antigenic stimulus has disappeared. Activating mutations in *CARD11* (which encodes Carma-1) have been detected in activated B cell-like diffuse large B cell lymphoma, another B cell malignancy^{76,77}. Such mutations generate constitutively active Carma-1 that associates with the Bcl-10-MALT1 complex without antigenic stimulation, which results in persistent activation of NF- κ B⁷⁸. A mutation that modifies the TLR adaptor MyD88 and promotes constitutive TLR signaling has been found in diffuse large B cell lymphoma of the activated B cell-like type. The L265P MyD88 variant promotes lymphoma cell survival by spontaneously assembling a protein complex containing the IRAK1 and IRAK4, leading to kinase activity of IRAK4, phosphorylation of IRAK1, signaling by NF- κ B, activation of the transcription factor STAT3 mediated by the kinase Jak, and secretion of IL-6, IL-10 and interferon- β ⁷⁹. Other mutations that lead to constitutive activation of the kinase NIK and result in the activation of both classical and alternative NF- κ B signaling have been detected in multiple myeloma, another type of B cell malignancy^{78,80-82}. Multiple myeloma-associated mutations include those in *NFKB2*, *BTRC*, *CARD11*, *CYLD*, *IKBIP*, *IKBKB*, *MAP3K1*, *MAP3K14*, *RIPK4*, *TLR4*, *TNFRSF1A*, *BIRC2*, *BIRC3*, *TRAF2* and *TRAF3*. *BTRC* encodes β -TrCP, the substrate-recognition subunit of the I κ B ubiquitin ligase⁸³. Products of the *BIRC2*, *BIRC3*, *TRAF2* and *TRAF3* loci form a ubiquitin-ligase complex responsible for degradative, Lys48-linked ubiquitination of NIK that keeps the concentration of this kinase below a critical threshold required for its autoactivation; and mutations and deletions of these genes affect this process. Other multiple myeloma-linked mutations have been found in *NIK* itself that affect the binding site for TRAF3, which connects NIK to the complex of TRAF2 and the ubiquitin ligases cIAP1 and cIAP2 (refs. 80,84,85). These mutations cause the inhibition of NIK turnover, which results in its autoactivation and the subsequent phosphorylation of IKK α , the key kinase responsible for activation of NF- κ B2 processing and the generation of alternative dimers of the p52 and RelB subunits of NF- κ B^{85,86}. NIK can also be overexpressed as a result of gene amplification or chromosomal translocations, which also occur in multiple myeloma⁷⁸.

By analogy to B cell lymphomas in which NF- κ B2 is constitutively processed as the result of its truncation because of chromosomal translocation⁷⁰, multiple myeloma has been proposed to depend on NIK-driven NF- κ B2 activation^{80,81}. Given those expectations, it was rather surprising when NIK-driven IKK β activation turned out to be more important for the survival of multiple myeloma cells than is NIK-driven activation of IKK α ⁸⁷.

The bulk of NF- κ B-positive tumors, however, are solid malignancies derived mainly from epithelial cells. NF- κ B-activating mutations are extremely rare in carcinomas, although mutations and gene fusions of *IKKA* and *IKKB* have been detected through genomic sequencing of breast and prostate cancer, respectively^{88,89}. In addition, the IKK-like kinase IKK ϵ has been identified as a contributor to the malignant activity of breast carcinoma cells⁹⁰. However, the role of IKK ϵ in the activation of NF- κ B is not well established and, therefore, its oncogenic activity may be NF- κ B independent. A role for IKK α in the self-renewal of breast cancer progenitors has been demonstrated in a mouse model⁹¹, and IKK α has been shown to be responsible for the tumor-promoting effect of progesterone in breast cancer, which is mediated through induction of the IKK α -activating cytokine RANKL in mammary epithelial cells^{92,93}. IKK α activation is also important for the metastatic spread of breast cancer; this depends on the production of RANKL, which in advanced and progesterone-independent tumors is produced by tumor-infiltrating regulatory T cells rather than carcinoma cells⁹⁴. NF- κ B activation is also involved in the *in vitro* formation of breast cancer stem cells in response to activation of the Src tyrosine kinase⁹⁵. However, the oncogenic functions of IKK α in breast or prostate cancers^{96,97} are not mediated through either classical or alternative NF- κ B signaling and instead depend on the nuclear functions of IKK α ⁹⁷. Notably, in the bulk of carcinomas in which classical NF- κ B signaling is activated and may provide the cancer cell with a survival advantage, the actual cause of NF- κ B activation remains to be identified and is probably microenvironmental factors rather than genetic alterations.

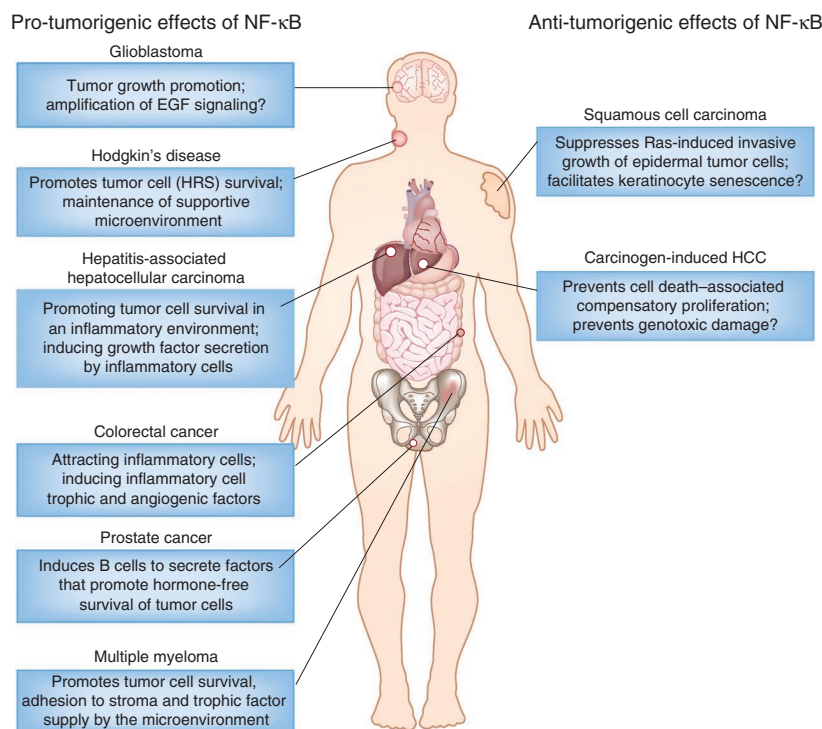


Figure 3 Pro- and anti-tumorigenic effects of NF-κB activation in cancer cells and their microenvironment. Opposing NF-κB inhibition effects are found in distinct cancer types, yet also in cancers of a similar type, depending on the mechanism of carcinogenesis. Hence, whereas NF-κB inhibition suppresses inflammation (hepatitis)-associated liver cancer (HCC), it facilitates carcinogen-induced HCC.

agents on hepatocarcinogenesis are yet to be demonstrated. On the basis of analysis of a CAC model in which tumor initiation depends on metabolic activation of the mutagen azoxymethane (AOM), it was concluded that NF-κB activation promotes tumor development after the initiation stage, most probably during early tumor promotion^{71,74}. Although the actual tumor-initiation event in *Mdr2*^{-/-} mice is not known, NF-κB probably also acts during the tumor-promotion stage by protecting premalignant cells from apoptotic elimination. Another important mechanism through which NF-κB contributes to tumor promotion as well as tumor progression is enhanced cell proliferation. At least in CAC, in which this aspect has been investigated in some detail, the

proliferative function of NF-κB is indirect and is mediated through IL-6 and related cytokines produced by myeloid cells that lead to the activation of STAT3 in IECs^{55,99}. Ablation of STAT3 in IECs also inhibits CAC development, affecting both cell survival and cell proliferation⁹⁹.

The relationships between NF-κB and STAT3 are complex (Fig. 2). In many cell types and circumstances, NF-κB and STAT3 control the expression of a similar repertoire of antiapoptotic genes^{102,103}. NF-κB and STAT3 can both interfere with synthesis of the tumor suppressor p53 and attenuate p53-mediated genomic surveillance¹⁰⁴. STAT3 controls the expression of c-Myc and cyclin D^{103,105}. Although NF-κB may control expression of those pro-proliferative factors in some cell types, ablation of IKKβ in IECs has no effect on cell proliferation⁵⁵, and in hepatocytes initiated by diethylnitrosamine (DEN), inhibition of NF-κB enhances cyclin D expression and cell proliferation¹⁰⁶. This seemingly paradoxical effect is probably due to the activation of STAT3 in IKKβ-depleted HCC cells¹⁰⁷, an outcome of NF-κB inhibition that is also observed in neutrophils⁶⁰. However, in other tumor types, NF-κB potentiates STAT3-mediated transactivation of genes encoding distinct inflammatory and pro-proliferative molecules in cells of the immune system present in the tumor microenvironment¹⁰³. Furthermore non-phosphorylated STAT3 is reported to activate the transcription of genes encoding cytokines and growth-promoting molecules via NF-κB¹⁰⁸.

The microenvironmental functions of NF-κB are widespread and complex. In addition to promoting the expression of inflammatory cytokines, NF-κB seems to be involved in the polarization of tumor-associated macrophages¹⁰⁹. Inhibition of NF-κB in such cells converts them from the M2 tumor-promoting phenotype to the M1 cytotoxic phenotype, thereby augmenting tumor regression¹¹⁰. Interestingly, the p50 subunit of NF-κB is a key regulator of M2-driven inflammatory reactions *in vitro* and *in vivo*. It has been shown that p50 inhibits NF-κB-driven M1 polarization, and p50-deficient mice have exacerbated M1-driven inflammation and a defective ability to mount allergy- and helminth-driven M2-polarized inflammatory reactions¹¹¹. NF-κB also acts in cancer-associated fibroblasts, in which it promotes the expression of a proinflammatory gene signature, important for

The role of NF-κB in the tumor microenvironment

Notably, the presence of activated NF-κB in a tumor is not necessarily causal, and even when it is of importance, just like in inflammation, NF-κB can influence tumor development and progression both positively and negatively (Fig. 3). The first two examples of a critical positive role for NF-κB in linking inflammation with tumor development were colitis-associated colon cancer (CAC) and hepatitis-associated liver cancer^{55,98}. In CAC, a classical inflammation-driven cancer that accounts for about 5% of sporadic colorectal cancers, it has been shown by conditional ablation of IKKβ that the activation of NF-κB in IECs, in which β-catenin signaling has been activated via mutation, provides premalignant progenitors with a survival advantage through the induction of antiapoptotic genes, such as that encoding Bcl-x_L (ref. 55). NF-κB in myeloid cells, most probably lamina propria macrophages, also makes an important contribution to tumor growth and progression through the transcriptional activation of genes encoding growth factors that enhance the proliferation of premalignant IECs and their transformed derivatives⁵⁵. Many inflammatory cytokines, including TNF, IL-6 and IL-23, produced by lamina propria macrophages and dendritic cells, as well as by tumor-associated macrophages, have been identified as the main drivers of CAC growth^{99–101}. Although TNF activates NF-κB in IECs and other epithelial cells, it should be noted that ablation of IKKβ in myeloid cells, which prevents TNF production, does not affect NF-κB or the survival of premalignant IECs⁵⁵. Thus, the actual cause of NF-κB activation in IECs remains to be identified. TNF produced by activated liver inflammatory cells, however, is probably responsible for NF-κB activation in hepatocytes of *Mdr2*^{-/-} mice, which experience chronic low-grade inflammation caused by phospholipid accumulation due to absence of the MDR2 phospholipid pump⁹⁸. Inhibition of hepatocyte NF-κB through expression of a nondegradable variant of IκBα blocks the development of hepatocellular carcinoma (HCC) in *Mdr2*^{-/-} mice and enhances the apoptosis of premalignant hepatocytes⁹⁸. Similar results have been obtained by the administration of nonsteroidal anti-inflammatory drugs (NSAIDs) or anti-TNF drugs to *Mdr2*^{-/-} mice⁹⁸, yet the effects of long-term treatment with these

macrophage recruitment, neovascularization and tumor growth¹¹². Although some of those activities are mediated through inflammatory cytokines, others are mediated via chemokine expression. NF- κ B activation in cancer cells can also lead to the upregulation of chemokines that initiate and maintain the tumor microenvironment through the recruitment of immune-response and inflammatory cells, as well as of progenitors of cancer-associated fibroblasts³. In addition, NF- κ B can affect epithelial-to-mesenchymal transition through induction of the transcription factors *twist* and *snail*^{113,114}, but these effects need to be confirmed *in vivo*. In summary, NF- κ B is involved in most if not all aspects of tumorigenesis, and many of its important activities are exerted in the tumor microenvironment.

Antitumorigenic effects of NF- κ B

The role of NF- κ B in cancer is not always positive (Fig. 3). Thus, blockade of NF- κ B via overexpression of $\text{I}\kappa\text{B}\alpha$ promotes oncogenic Ras-induced invasive epidermal growth resembling squamous cell carcinoma¹¹⁵. Although the mechanism of this phenomenon is not obvious, it might be related to the role of NF- κ B in oncogene-induced senescence¹¹⁶; blocking NF- κ B might abolish Ras-induced senescence. In mice given DEN, hepatocyte-specific ablation of IKK β strongly enhances the development of HCC¹⁰⁶. A similar enhancement of DEN-induced HCC development has been found after hepatocyte-specific ablation of the inflammation-responsive protein kinase p38 α ^{117,118}. Although IKK β and p38 α do not control a common set of target genes, they both maintain hepatocyte viability by suppressing ROS accumulation¹¹⁸. Indeed, the ability of IKK β and p38 α to suppress HCC development is related to their ability to prevent DEN-induced cell death that otherwise triggers compensatory proliferation, which is critical for the transmission of oncogenic mutations, as mature hepatocytes with such mutations do not proliferate unless the liver is damaged. Hepatocyte-specific ablation of IKK γ ¹⁹ or TAK1 (refs. 120,121), which are both required for the activation of IKK and NF- κ B, results in spontaneous liver damage, hepatocyte death, liver fibrosis and spontaneous development of HCC. In this case, however, the cause of the oncogenic mutations that are propagated via compensatory proliferation is unknown. Liver damage and subsequent HCC development in mice lacking hepatocyte IKK γ is related to ROS accumulation, as both can be prevented by administration of the potent antioxidant butylated hydroxyanisole, which has been found to suppress the enhanced HCC development in mice lacking hepatocyte IKK β ¹⁰⁶. One further clue to the putative antitumorigenic activity of NF- κ B is provided by a mouse model of stomach-specific overexpression of IL-1 β , whose endogenous production is negatively regulated by NF- κ B^{59,60}. Mice with transgenic expression of IL-1 β develop gastric inflammation and cancer, possibly due to the recruitment of myeloid-derived suppressor cells to the stomach¹²².

The tumor-suppressive function of hepatocyte NF- κ B applies only to situations in which the main driver of liver inflammation is hepatocyte death, which results in release of IL-1 α , thereby triggering pro-tumorigenic NF- κ B signaling in Kupffer cells¹¹⁸. In another model of chronic liver inflammation that depends on NF- κ B activation in hepatocytes, this one driven by ectopic expression of the TNF family member lymphotoxin, ablation of hepatocyte IKK β prevents HCC development¹²³. Furthermore, even in the DEN model, ablation of IKK β in Kupffer cells inhibits HCC development¹⁰⁶, which is also inhibited by ablation of the IL-1 receptor and MyD88, both of which are required for activation of NF- κ B in Kupffer cells¹¹⁸. Activated Kupffer cells produce the critical tumor-promoting cytokine IL-6, whose ablation almost completely prevents the induction of HCC by

DEN¹²⁴. As in CAC, IL-6 acts via STAT3, whose hepatocyte-specific ablation also inhibits DEN-induced HCC¹⁰⁷. Ablation of IKK β causes STAT3 activation as a result of enhanced ROS accumulation, and inverse relationships between the activation of NF- κ B and STAT3 have also been observed in human HCC¹⁰⁷. IKK β ablation also enhances the activation of Jnk family members¹⁰⁶, including Jnk1, which contributes to HCC development¹¹⁸ (Fig. 2).

Another interesting example of a cell type-specific role for NF- κ B in tumor progression is castration-resistant prostate cancer⁹⁶. In this case, it has been found that IKK β activation in B cells is required for the production of lymphotoxin composed of two subunits encoded by target genes of NF- κ B; this lymphotoxin activates IKK α in prostate carcinoma cells. Whereas IKK β ablation in prostate carcinoma cells has no effect on the development or recurrence of tumors, ablation or inactivation of IKK α delays or inhibits the development of castration-resistant cancer⁹⁶. Furthermore, IKK α , not IKK β , is required for the metastatic spread of prostate cancer in mice⁹⁷. It should be noted, however, that this pro-metastatic function of IKK α is NF- κ B independent and requires nuclear translocation of IKK α .

Blocking NF- κ B for cancer prevention and therapy

It is conceivable that if indications for NF- κ B inhibition as means of cancer treatment had to be prioritized, such a list would be headed by tumors bearing NF- κ B-activating mutations⁷⁸, followed by tumors in which NF- κ B activation is linked to a bad prognosis¹²⁵ and those with NF- κ B activation due to microenvironmental factors¹²⁶. For some time, the first group consisted almost completely of hematological malignancies, lymphomas, leukemias and multiple myeloma, in which a variety of mutations in genes encoding components of the NF- κ B pathway have been found^{79,81,127}, yet deletions of $\text{I}\kappa\text{B}\alpha$ have now also been identified in brain tumors³⁸. When treatment for such cancers is considered, the strategy should be tailored to the patient, depending on the nature of the activating mutation; hence, it would not be wise to use IKK inhibitors to treat Hodgkin's disease and glioblastoma tumors that have deletion of $\text{I}\kappa\text{B}\alpha$, as the main substrate of those reagents is missing, but RelA-specific NF- κ B decoys⁵¹ would be a logical choice. It should be noted, however, that even tumors bearing activating mutations of the gene encoding NF- κ B in cancer-initiating stem cells, such as multiple myeloma, diffuse large B cell lymphoma and Hodgkin's lymphoma, often also benefit from NF- κ B activation in the microenvironment¹¹⁴. Thus, blocking the activation of NF- κ B in the microenvironment may compromise tumorigenesis regardless of the activating mutation; it may even be advantageous because of the relative genomic stability of the cells in the surrounding microenvironment and their likely lower tendency to develop drug resistance.

Which therapeutic means are available clinically or experimentally for targeting NF- κ B oncogenic pathways? To our best knowledge, although many IKK inhibitors have been developed and have been found to exert antitumor effects in a variety of experimental cancer models, ranging from lymphoma to melanoma^{128–131}, at present no such drug has been clinically approved. The fairly limited success of IKK inhibitors, like that of many other targeted cancer therapeutics when they are used as single agents, has prompted studies seeking to stratify failure versus success or combining targeted therapeutics with traditional chemotherapy. IKK inhibitors, for example, should be effective in sensitizing cancer cells to standard chemotherapy-induced death, given the suppression of NF- κ B-dependent genes encoding antiapoptotic molecules¹³² and antioxidant molecules (such as ferritin heavy chain)¹³³. An alternative strategy that has attracted growing interest is a new class of compounds known as multitarget drugs¹³⁴.

These compounds were selected to improve therapeutic efficacy by targeting diverse regulatory pathways essential for the proliferation and survival of cancer cells. Data-driven computational modeling techniques aim to find key vectors that represent signal combinations that contain information necessary for the prediction of cell responses to various perturbations¹³⁴. Among the promising roads located by this approach is a combination of blockers of the heat-shock protein hsp90 and inhibitors of histone deacetylases and the ubiquitin-proteasome system^{135–137}. Combinations of bortezomib, a proteasome inhibitor that can block NF- κ B activation, and various inhibitors of histone deacetylase have already found their way into advanced clinical studies. The most extensively studied is a combination of bortezomib and vorinostat (inhibitors of histone deacetylase). A phase III trial of this pair is being conducted in patients with relapsed and/or refractory multiple myeloma after a very good response in a third of the patients who failed to respond to bortezomib monotherapy (US National Institutes of Health Clinical Trials identifier NCT00773747)¹³⁸. An interesting new addition to the arsenal of ubiquitin-proteasome-system inhibitors is MLN4924, which inhibits the NEDD8-activating enzyme and blocks NF- κ B signaling in primary diffuse large B cell lymphoma, resulting in tumor regression¹³⁹. NEDD8-activating enzyme facilitates the addition of the ubiquitin-like protein NEDD8 to Cul-1; this is required for activity of the Skp1–Cul1–F-box complex SCF β -TrCP, an I κ B–E3 ubiquitin ligase, which promotes several NF- κ B signaling steps¹⁴⁰. By itself, the F-box protein β -TrCP is a likely target for NF- κ B inhibition¹⁴¹, yet no effective small molecules targeting this enzyme have been reported.

Although there is a rationale for inhibiting NF- κ B in tumors with constitutive or chemotherapy-induced NF- κ B activation, caution should be taken with other types of cancer in which NF- κ B activation could be a homeostatic switch, possibly limiting genotoxic damage¹⁴² or toning down an inflated innate immune response⁵⁸. Such cases, so far apparent only in experimental cancer systems, should be taken into account when an NF- κ B-blocking therapeutic regimen is being considered, and this should possibly be handled with a drug combination that will rebalance the adverse effects of NF- κ B inhibition. Thus, if inhibition of NF- κ B promotes enhanced secretion of IL-1 β and neutrophilia⁶⁰, anti-IL-1 therapy may reverse this possible pro-tumorigenic effect, albeit at the high cost of greater susceptibility to infection.

Another issue is prevention and prophylactic therapy. Will long-term suppression of smoldering inflammation result in a lower cancer risk? As NF- κ B inhibitors have not yet entered clinical practice, the information available is limited to the long-term effects of NSAIDs, mostly aspirin. Daily aspirin use at doses as low as 75 mg per day for 5 years or longer has been found to diminish death due to several common cancers, with 55–75% lower risk of death for the main types of cancer, such as colorectal, pancreatic and lung carcinomas¹⁴³. For colorectal cancer, this effect is probably not, as previously thought, due to selective inhibition of cyclooxygenase 2 (COX-2), as adjuvant therapy with the selective COX-2 inhibitor rofecoxib does not improve overall survival, nor does COX-2 expression correlate with prognosis nor can it be used to predict the effectiveness of COX-2 inhibitors¹⁴⁴. Thus, although the specific target of NSAID-based cancer chemoprevention remains unknown, its remarkable preventive effect suggests that low-grade inflammation may be a far more important factor than previously appreciated. It has been suggested that 20% of all cancers are linked to inflammation¹⁴⁵, but this figure is probably actually much higher¹⁴⁶. Systems biology-type studies should be helpful in identifying critical nodes of inflammation signaling for drug targeting. Nevertheless, it is probably safe to predict that

NF- κ B will emerge as an important hub in any inflammation network, which should motivate the development of effective inhibitors of the NF- κ B pathway, particularly those that evade the proinflammatory side effects discussed above. NF- κ B-targeting drugs may eventually prove more effective and possibly safer than NSAID use for cancer chemoprevention and therapy.

Concluding remarks

Tumor-promoting inflammation has finally been recognized as one of the hallmarks of cancer¹⁴⁷. Carcinogenesis is a multistage process, and although in the classical sequence of chemical carcinogenesis, tumor initiation is followed by tumor promotion¹⁴⁸, inflammation represents an inverse carcinogenesis program: tumor-promoting inflammation may precede tumor initiation, creating a favorable microenvironment in which cells with cancer-causing mutations thrive. Human epidemiology and animal model studies indicate that chronic, smoldering inflammation may be a far more widespread ground for cancer development than previously thought, and NF- κ B activation, as one of the pillars of inflammation, may have a promoting role in most cancers. Twenty-five years after the discovery of NF- κ B, much (and yet not enough) has been learned about its signaling and transcriptional targets, and taming NF- κ B activity will remain an important challenge for modern cancer biology.

ACKNOWLEDGMENTS

We thank E. Pikarsky, I. Alkalay-Snir and A. Pribluda for comments and discussions. Supported by the Israel Science Foundation, Israel Cancer Research Fund, the Crohn's & Colitis Foundation of America, the German-Israeli Foundation, Dr. Miriam and Sheldon G. Adelson Medical Research Foundation, the US National Institutes of Health and the American Cancer Society.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

Published online at <http://www.nature.com/natureimmunology/>.

Reprints and permissions information is available online at <http://www.nature.com/reprints/index.html>.

- Balkwill, F. & Mantovani, A. Cancer and inflammation: implications for pharmacology and therapeutics. *Clin. Pharmacol. Ther.* **87**, 401–406 (2010).
- Biswas, S.K. & Mantovani, A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat. Immunol.* **11**, 889–896 (2010).
- Grivennikov, S.I., Greten, F.R. & Karin, M. Immunity, inflammation, and cancer. *Cell* **140**, 883–899 (2010).
- Erez, N. & Coussens, L.M. Leukocytes as paracrine regulators of metastasis and determinants of organ-specific colonization. *Int. J. Cancer* **128**, 2536–2544 (2011).
- Qian, B.Z. & Pollard, J.W. Macrophage diversity enhances tumor progression and metastasis. *Cell* **141**, 39–51 (2010).
- Medzhitov, R. Inflammation 2010: new adventures of an old flame. *Cell* **140**, 771–776 (2010).
- Muller, W.E., Korzhnev, M., Le Pennec, G., Muller, I.M. & Schroder, H.C. Origin of metazoan stem cell system in sponges: first approach to establish the model (*Suberites domuncula*). *Biomol. Eng.* **20**, 369–379 (2003).
- Barnes, P.J. & Karin, M. Nuclear factor- κ B: a pivotal transcription factor in chronic inflammatory diseases. *N. Engl. J. Med.* **336**, 1066–1071 (1997).
- Tak, P.P. & Firestein, G.S. NF- κ B: a key role in inflammatory diseases. *J. Clin. Invest.* **107**, 7–11 (2001).
- Foxwell, B.M., Bondeson, J., Brennan, F. & Feldmann, M. Adenoviral transgene delivery provides an approach to identifying important molecular processes in inflammation: evidence for heterogeneity in the requirement for NF κ B in tumour necrosis factor production. *Ann. Rheum. Dis.* **59** (Suppl 1), i54–i59 (2000).
- Lawrence, T., Gilroy, D.W., Colville-Nash, P.R. & Willoughby, D.A. Possible new role for NF- κ B in the resolution of inflammation. *Nat. Med.* **7**, 1291–1297 (2001).
- Augustin, R., Fraune, S. & Bosch, T.C. How Hydra senses and destroys microbes. *Semin. Immunol.* **22**, 54–58 (2010).
- Hemmrich, G., Miller, D.J. & Bosch, T.C. The evolution of immunity: a low-life perspective. *Trends Immunol.* **28**, 449–454 (2007).
- Lange, C. *et al.* Defining the origins of the NOD-like receptor system at the base of animal evolution. *Mol. Biol. Evol.* **28**, 1687–1702 (2007).
- Srivastava, M. *et al.* The *Amphimedon queenslandica* genome and the evolution of animal complexity. *Nature* **466**, 720–726 (2010).

16. Sarkar, D., Desalle, R. & Fisher, P.B. Evolution of MDA-5/RIG-I-dependent innate immunity: independent evolution by domain grafting. *Proc. Natl. Acad. Sci. USA* **105**, 17040–17045 (2008).
17. Domazet-Loso, T. & Tautz, D. Phylostratigraphic tracking of cancer genes suggests a link to the emergence of multicellularity in metazoa. *BMC Biol.* **8**, 66 (2010).
18. Guttridge, D.C., Albanese, C., Reuther, J.Y., Pestell, R.G. & Baldwin, A.S. Jr. NF- κ B controls cell growth and differentiation through transcriptional regulation of cyclin D1. *Mol. Cell. Biol.* **19**, 5785–5799 (1999).
19. La Rosa, F.A., Pierce, J.W. & Sonenshein, G.E. Differential regulation of the c-myc oncogene promoter by the NF- κ B rel family of transcription factors. *Mol. Cell. Biol.* **14**, 1039–1044 (1994).
20. Duckett, C.S. Apoptosis and NF- κ B: the FADD connection. *J. Clin. Invest.* **109**, 579–580 (2002).
21. Dhawan, S., Singh, S. & Aggarwal, B.B. Induction of endothelial cell surface adhesion molecules by tumor necrosis factor is blocked by protein tyrosine phosphatase inhibitors: role of the nuclear transcription factor NF- κ B. *Eur. J. Immunol.* **27**, 2172–2179 (1997).
22. Collins, T. *et al.* Transcriptional regulation of endothelial cell adhesion molecules: NF- κ B and cytokine-inducible enhancers. *FASEB J.* **9**, 899–909 (1995).
23. Cao, Y. & Karin, M. NF- κ B in mammary gland development and breast cancer. *J. Mammary Gland Biol. Neoplasia* **8**, 215–223 (2003).
24. Snapper, C.M. *et al.* B cells from p50/NF- κ B knockout mice have selective defects in proliferation, differentiation, germ-line CH transcription, and Ig class switching. *J. Immunol.* **156**, 183–191 (1996).
25. Nickols, J.C., Valentine, W., Kanwal, S. & Carter, B.D. Activation of the transcription factor NF- κ B in Schwann cells is required for peripheral myelin formation. *Nat. Neurosci.* **6**, 161–167 (2003).
26. Cordero, J.B. *et al.* Oncogenic Ras diverts a host TNF tumor suppressor activity into tumor promoter. *Dev. Cell* **18**, 999–1011 (2010).
27. Feng, Y., Santoriello, C., Mione, M., Hurlstone, A. & Martin, P. Live imaging of innate immune cell sensing of transformed cells in zebrafish larvae: parallels between tumor initiation and wound inflammation. *PLoS Biol.* **8**, e1000562 (2010).
28. Squires, D.F. Neoplasia in a coral? *Science* **148**, 503–505 (1965).
29. Wiebecke, B., Brandts, A. & Eder, M. Epithelial proliferation and morphogenesis of hyperplastic adenomatous and villous polyps of the human colon. *Virchows Arch. A Pathol. Anat. Histol.* **364**, 35–49 (1974).
30. Cole, J.W. & McKalen, A. Studies on the morphogenesis of adenomatous polyps in the human Colon. *Cancer* **16**, 998–1002 (1963).
31. Dvorak, H.F. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N. Engl. J. Med.* **315**, 1650–1659 (1986).
32. Eaves, C.J. Cancer stem cells: Here, there, everywhere? *Nature* **456**, 581–582 (2008).
33. Shackleton, M., Quintana, E., Fearon, E.R. & Morrison, S.J. Heterogeneity in cancer: cancer stem cells versus clonal evolution. *Cell* **138**, 822–829 (2009).
34. Barker, N. *et al.* Crypt stem cells as the cells-of-origin of intestinal cancer. *Nature* **457**, 608–611 (2009).
35. Sato, T. *et al.* Paneth cells constitute the niche for Lgr5 stem cells in intestinal crypts. *Nature* **469**, 415–418 (2010).
36. Apidianakis, Y., Pitsouli, C., Perrimon, N. & Rahme, L. Synergy between bacterial infection and genetic predisposition in intestinal dysplasia. *Proc. Natl. Acad. Sci. USA* **106**, 20883–20888 (2009).
37. Foster, S.L., Hargreaves, D.C. & Medzhitov, R. Gene-specific control of inflammation by TLR-induced chromatin modifications. *Nature* **447**, 972–978 (2007).
38. Bredel, M. *et al.* NF- κ B deletion in glioblastomas. *N. Engl. J. Med.* **364**, 627–637 (2010).
39. Bivona, T.G. *et al.* FAS and NF- κ B signalling modulate dependence of lung cancers on mutant EGFR. *Nature* **471**, 523–526 (2011).
40. Smale, S. Hierarchies of NF- κ B target-gene regulation. *Nat. Immunol.* **12**, 689–694 (2011).
41. Bonizzi, G. & Karin, M. The two NF- κ B activation pathways and their role in innate and adaptive immunity. *Trends Immunol.* **25**, 280–288 (2004).
42. Gewirtz, A.T. *et al.* Salmonella typhimurium induces epithelial IL-8 expression via Ca(2+)-mediated activation of the NF- κ B pathway. *J. Clin. Invest.* **105**, 79–92 (2000).
43. Lavon, I. *et al.* High susceptibility to bacterial infection, but no liver dysfunction, in mice compromised for hepatocyte NF- κ B activation. *Nat. Med.* **6**, 573–577 (2000).
44. Kaser, A., Zeissig, S. & Blumberg, R.S. Inflammatory bowel disease. *Annu. Rev. Immunol.* **28**, 573–621 (2010).
45. Neurath, M.F., Pettersson, S., Meyer zum Buschenfelde, K.H. & Strober, W. Local administration of antisense phosphorothioate oligonucleotides to the p65 subunit of NF- κ B abrogates established experimental colitis in mice. *Nat. Med.* **2**, 998–1004 (1996).
46. Tak, P.P. *et al.* Inhibitor of nuclear factor κ B kinase β is a key regulator of synovial inflammation. *Arthritis Rheum.* **44**, 1897–1907 (2001).
47. Lizzul, P.F. *et al.* Differential expression of phosphorylated NF- κ B/RelA in normal and psoriatic epidermis and downregulation of NF- κ B in response to treatment with etanercept. *J. Invest. Dermatol.* **124**, 1275–1283 (2005).
48. Williams, R.O., Paleolog, E. & Feldmann, M. Cytokine inhibitors in rheumatoid arthritis and other autoimmune diseases. *Curr. Opin. Pharmacol.* **7**, 412–417 (2007).
49. MacMaster, J.F. *et al.* An inhibitor of I κ B kinase, BMS-345541, blocks endothelial cell adhesion molecule expression and reduces the severity of dextran sulfate sodium-induced colitis in mice. *Inflamm. Res.* **52**, 508–511 (2003).
50. Gillooly, K.M. *et al.* Periodic, partial inhibition of I κ B kinase β -mediated signaling yields therapeutic benefit in preclinical models of rheumatoid arthritis. *J. Pharmacol. Exp. Ther.* **331**, 349–360 (2009).
51. Miagkov, A.V. *et al.* NF- κ B activation provides the potential link between inflammation and hyperplasia in the arthritic joint. *Proc. Natl. Acad. Sci. USA* **95**, 13859–13864 (1998).
52. Schopf, L. *et al.* IKK β inhibition protects against bone and cartilage destruction in a rat model of rheumatoid arthritis. *Arthritis Rheum.* **54**, 3163–3173 (2006).
53. Pitts, W.J., Kempson, J. & John, E.M. (ed. Macor, J.E.) in *Annual Reports in Medicinal Chemistry*, Vol. 43, 155–170 (Academic, 2008).
54. Bohrer, H. *et al.* Role of NF κ B in the mortality of sepsis. *J. Clin. Invest.* **100**, 972–985 (1997).
55. Greten, F.R. *et al.* IKK β links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* **118**, 285–296 (2004).
56. Nenci, A. *et al.* Epithelial NEMO links innate immunity to chronic intestinal inflammation. *Nature* **446**, 557–561 (2007).
57. Pasparakis, M. Regulation of tissue homeostasis by NF- κ B signalling: implications for inflammatory diseases. *Nat. Rev. Immunol.* **9**, 778–788 (2009).
58. Greten, F.R. *et al.* NF- κ B is a negative regulator of IL-1 β secretion as revealed by genetic and pharmacological inhibition of IKK β . *Cell* **130**, 918–931 (2007).
59. Bruey, J.M. *et al.* Bcl-2 and Bcl-XL regulate proinflammatory caspase-1 activation by interaction with NALP1. *Cell* **129**, 45–56 (2007).
60. Hsu, L.C. *et al.* IL-1 β -driven neutrophilia preserves antibacterial defense in the absence of the kinase IKK β . *Nat. Immunol.* **12**, 144–150 (2011).
61. Genovese, M.C. *et al.* Combination therapy with etanercept and anakinra in the treatment of patients with rheumatoid arthritis who have been treated unsuccessfully with methotrexate. *Arthritis Rheum.* **50**, 1412–1419 (2004).
62. Apte, R.N. *et al.* Effects of micro-environment- and malignant cell-derived interleukin-1 in carcinogenesis, tumour invasiveness and tumour-host interactions. *Eur. J. Cancer* **42**, 751–759 (2006).
63. Ruland, J. Return to homeostasis—downregulation of NF- κ B responses. *Nat. Immunol.* **12**, 709–714 (2011).
64. O’Dea, E. & Hoffmann, A. The regulatory logic of the NF- κ B signaling system. *Cold Spring Harb. Perspect. Biol.* **2**, a000216 (2010).
65. Chen, L.W. *et al.* The two faces of IKK and NF- κ B inhibition: prevention of systemic inflammation but increased local injury following intestinal ischemia-reperfusion. *Nat. Med.* **9**, 575–581 (2003).
66. Eckmann, L. *et al.* Opposing functions of IKK β during acute and chronic intestinal inflammation. *Proc. Natl. Acad. Sci. USA* **105**, 15058–15063 (2008).
67. Gilmore, T.D. The Rel1/NF- κ B/I κ B signal transduction pathway and cancer. *Cancer Treat. Res.* **115**, 241–265 (2003).
68. Cabanes, A. *et al.* Enhancement of antitumor activity of polyethylene glycol-coated liposomal doxorubicin with soluble and liposomal interleukin 2. *Clin. Cancer Res.* **5**, 687–693 (1999).
69. Franzoso, G. *et al.* The candidate oncoprotein Bcl-3 is an antagonist of p50/NF- κ B-mediated inhibition. *Nature* **359**, 339–342 (1992).
70. Neri, A. *et al.* B cell lymphoma-associated chromosomal translocation involves candidate oncogene *lyt-10*, homologous to NF- κ B p50. *Cell* **67**, 1075–1087 (1991).
71. Karin, M., Cao, Y., Greten, F.R. & Li, Z.W. NF- κ B in cancer: from innocent bystander to major culprit. *Nat. Rev. Cancer* **2**, 301–310 (2002).
72. Willis, T.G. *et al.* Bcl10 is involved in t(1;14)(p22;q32) of MALT B cell lymphoma and mutated in multiple tumor types. *Cell* **96**, 35–45 (1999).
73. Uren, A.G. *et al.* Identification of paracaspases and metacaspases: two ancient families of caspase-like proteins, one of which plays a key role in MALT lymphoma. *Mol. Cell* **6**, 961–967 (2000).
74. Hacker, H. & Karin, M. Regulation and function of IKK and IKK-related kinases. *Sci. STKE* **2006**, re13 (2006).
75. Wertz, I.E. & Dixit, V.M. Signaling to NF- κ B: regulation by ubiquitination. *Cold Spring Harb. Perspect. Biol.* **2**, a003350 (2010).
76. Lenz, G. *et al.* Oncogenic CARD11 mutations in human diffuse large B cell lymphoma. *Science* **319**, 1676–1679 (2008).
77. Ngo, V.N. *et al.* A loss-of-function RNA interference screen for molecular targets in cancer. *Nature* **441**, 106–110 (2006).
78. Staudt, L.M. Oncogenic activation of NF- κ B. *Cold Spring Harb. Perspect. Biol.* **2**, a000109 (2010).
79. Ngo, V.N. *et al.* Oncogenically active MYD88 mutations in human lymphoma. *Nature* **470**, 115–119 (2011).
80. Annunziata, C.M. *et al.* Frequent engagement of the classical and alternative NF- κ B pathways by diverse genetic abnormalities in multiple myeloma. *Cancer Cell* **12**, 115–130 (2007).
81. Keats, J.J. *et al.* Promiscuous mutations activate the noncanonical NF- κ B pathway in multiple myeloma. *Cancer Cell* **12**, 131–144 (2007).
82. Chapman, M.A. *et al.* Initial genome sequencing and analysis of multiple myeloma. *Nature* **471**, 467–472 (2011).
83. Yaron, A. *et al.* Identification of the receptor component of the I κ B α -ubiquitin ligase. *Nature* **396**, 590–594 (1998).
84. Liao, G., Zhang, M., Harhaj, E.W. & Sun, S.C. Regulation of the NF- κ B-inducing kinase by tumor necrosis factor receptor-associated factor 3-induced degradation. *J. Biol. Chem.* **279**, 26243–26250 (2004).
85. Vallabhapurapu, S. & Karin, M. Regulation and function of NF- κ B transcription factors in the immune system. *Annu. Rev. Immunol.* **27**, 693–733 (2009).

86. Sasaki, Y. *et al.* NIK overexpression amplifies, whereas ablation of its TRAF3-binding domain replaces BAFF:BAFF-R-mediated survival signals in B cells. *Proc. Natl. Acad. Sci. USA* **105**, 10883–10888 (2008).
87. Lam, L.T. *et al.* Compensatory IKK α activation of classical NF- κ B signaling during IKK β inhibition identified by an RNA interference sensitization screen. *Proc. Natl. Acad. Sci. USA* **105**, 20798–20803 (2008).
88. Pflueger, D. *et al.* Discovery of non-ETS gene fusions in human prostate cancer using next-generation RNA sequencing. *Genome Res.* (2010).
89. Stratton, M.R., Campbell, P.J. & Futreal, P.A. The cancer genome. *Nature* **458**, 719–724 (2009).
90. Boehm, J.S. *et al.* Integrative genomic approaches identify IKBKE as a breast cancer oncogene. *Cell* **129**, 1065–1079 (2007).
91. Cao, Y., Luo, J.L. & Karin, M. I κ B kinase α kinase activity is required for self-renewal of ErbB2/Her2-transformed mammary tumor-initiating cells. *Proc. Natl. Acad. Sci. USA* **104**, 15852–15857 (2007).
92. Gonzalez-Suarez, E. *et al.* RANK ligand mediates progesterin-induced mammary epithelial proliferation and carcinogenesis. *Nature* **468**, 103–107 (2010).
93. Schramek, D. *et al.* Osteoclast differentiation factor RANKL controls development of progesterin-driven mammary cancer. *Nature* **468**, 98–102 (2010).
94. Tan, W. *et al.* Tumor-infiltrating T regulatory cells stimulate mammary cancer metastasis through RANKL-RANK signaling. *Nature* **470**, 548–553 (2011).
95. Iliopoulos, D., Hirsch, H.A. & Struhl, K. An epigenetic switch involving NF- κ B, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. *Cell* **139**, 693–706 (2009).
96. Ammirante, M., Luo, J.L., Grivennikov, S., Nedospasov, S. & Karin, M. B-cell-derived lymphotoxin promotes castration-resistant prostate cancer. *Nature* **464**, 302–305 (2010).
97. Luo, J.L. *et al.* Nuclear cytokine-activated IKK α controls prostate cancer metastasis by repressing Maspin. *Nature* **446**, 690–694 (2007).
98. Pikarsky, E. *et al.* NF- κ B functions as a tumour promoter in inflammation-associated cancer. *Nature* **431**, 461–466 (2004).
99. Grivennikov, S. *et al.* IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell* **15**, 103–113 (2009).
100. Popivanova, B.K. *et al.* Blocking TNF- α in mice reduces colorectal carcinogenesis associated with chronic colitis. *J. Clin. Invest.* **118**, 560–570 (2008).
101. Terzic, J., Grivennikov, S., Karin, E. & Karin, M. Inflammation and colon cancer. *Gastroenterology* **138**, 2101–2114 (2010).
102. Karin, M. Nuclear factor- κ B in cancer development and progression. *Nature* **441**, 431–436 (2006).
103. Yu, H., Kortylewski, M. & Pardoll, D. Crosstalk between cancer and immune cells: role of STAT3 in the tumour microenvironment. *Nat. Rev. Immunol.* **7**, 41–51 (2007).
104. Colotta, F., Allavena, P., Sica, A., Garlanda, C. & Mantovani, A. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* **30**, 1073–1081 (2009).
105. Bollrath, J. *et al.* gp130-mediated Stat3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. *Cancer Cell* **15**, 91–102 (2009).
106. Maeda, S., Kamata, H., Luo, J.L., Leffert, H. & Karin, M. IKK β couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell* **121**, 977–990 (2005).
107. He, G. *et al.* Hepatocyte IKK β /NF- κ B inhibits tumor promotion and progression by preventing oxidative stress-driven STAT3 activation. *Cancer Cell* **17**, 286–297 (2010).
108. Yang, J. *et al.* Unphosphorylated STAT3 accumulates in response to IL-6 and activates transcription by binding to NF κ B. *Genes Dev.* **21**, 1396–1408 (2007).
109. Mantovani, A. & Sica, A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr. Opin. Immunol.* **22**, 231–237 (2010).
110. Hagemann, T. *et al.* 'Re-educating' tumor-associated macrophages by targeting NF- κ B. *J. Exp. Med.* **205**, 1261–1268 (2008).
111. Porta, C. *et al.* Tolerance and M2 (alternative) macrophage polarization are related processes orchestrated by p50 nuclear factor κ B. *Proc. Natl. Acad. Sci. USA* **106**, 14978–14983 (2009).
112. Erez, N., Truitt, M., Olson, P., Arron, S.T. & Hanahan, D. Cancer-associated fibroblasts are activated in incipient neoplasia to orchestrate tumor-promoting inflammation in an NF- κ B-dependent manner. *Cancer Cell* **17**, 135–147 (2010).
113. Yu, H., Pardoll, D. & Jove, R. STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat. Rev. Cancer* **9**, 798–809 (2009).
114. Markovina, S. *et al.* Bone marrow stromal cells from multiple myeloma patients uniquely induce bortezomib resistant NF- κ B activity in myeloma cells. *Mol. Cancer* **9**, 176 (2010).
115. Dajee, M. *et al.* NF- κ B blockade and oncogenic Ras trigger invasive human epidermal neoplasia. *Nature* **421**, 639–643 (2003).
116. Acosta, J.C. *et al.* Chemokine signaling via the CXCR2 receptor reinforces senescence. *Cell* **133**, 1006–1018 (2008).
117. Hui, L. *et al.* p38 α suppresses normal and cancer cell proliferation by antagonizing the JNK-c-Jun pathway. *Nat. Genet.* **39**, 741–749 (2007).
118. Sakurai, T. *et al.* Hepatocyte necrosis induced by oxidative stress and IL-1 α release mediate carcinogen-induced compensatory proliferation and liver tumorigenesis. *Cancer Cell* **14**, 156–165 (2008).
119. Luedde, T. *et al.* Deletion of NEMO/IKKgamma in liver parenchymal cells causes steatohepatitis and hepatocellular carcinoma. *Cancer Cell* **11**, 119–132 (2007).
120. Inokuchi, S. *et al.* Disruption of TAK1 in hepatocytes causes hepatic injury, inflammation, fibrosis, and carcinogenesis. *Proc. Natl. Acad. Sci. USA* **107**, 844–849 (2010).
121. Bettermann, K. *et al.* TAK1 suppresses a NEMO-dependent but NF- κ B-independent pathway to liver cancer. *Cancer Cell* **17**, 481–496 (2010).
122. Tu, S. *et al.* Overexpression of interleukin-1 β induces gastric inflammation and cancer and mobilizes myeloid-derived suppressor cells in mice. *Cancer Cell* **14**, 408–419 (2008).
123. Haybaeck, J. *et al.* A lymphotoxin-driven pathway to hepatocellular carcinoma. *Cancer Cell* **16**, 295–308 (2009).
124. Naugler, W.E. *et al.* Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* **317**, 121–124 (2007).
125. Dunleavy, K. *et al.* Differential efficacy of bortezomib plus chemotherapy within molecular subtypes of diffuse large B-cell lymphoma. *Blood* **113**, 6069–6076 (2009).
126. Biswas, D.K. *et al.* NF- κ B activation in human breast cancer specimens and its role in cell proliferation and apoptosis. *Proc. Natl. Acad. Sci. USA* **101**, 10137–10142 (2004).
127. Compagno, M. *et al.* Mutations of multiple genes cause deregulation of NF- κ B in diffuse large B-cell lymphoma. *Nature* **459**, 717–721 (2009).
128. Amschler, K. *et al.* NF- κ B inhibition through proteasome inhibition or IKK β blockade increases the susceptibility of melanoma cells to cytostatic treatment through distinct pathways. *J. Invest. Dermatol.* **130**, 1073–1086 (2010).
129. Lam, L.T. *et al.* Small molecule inhibitors of I κ B kinase are selectively toxic for subgroups of diffuse large B-cell lymphoma defined by gene expression profiling. *Clin. Cancer Res.* **11**, 28–40 (2005).
130. Lee, D.F. & Hung, M.C. Advances in targeting IKK and IKK-related kinases for cancer therapy. *Clin. Cancer Res.* **14**, 5656–5662 (2008).
131. Schon, M. *et al.* KINK-1, a novel small-molecule inhibitor of IKK β , and the susceptibility of melanoma cells to antitumor treatment. *J. Natl. Cancer Inst.* **100**, 862–875 (2008).
132. Nakanishi, C. & Toi, M. Nuclear factor- κ B inhibitors as sensitizers to anticancer drugs. *Nat. Rev. Cancer* **5**, 297–309 (2005).
133. Kiessling, M.K. *et al.* Inhibition of constitutively activated nuclear factor- κ B induces reactive oxygen species- and iron-dependent cell death in cutaneous T-cell lymphoma. *Cancer Res.* **69**, 2365–2374 (2009).
134. Pritchard, J.R. *et al.* Three-kinase inhibitor combination recreates multipathway effects of a geldanamycin analogue on hepatocellular carcinoma cell death. *Mol. Cancer Ther.* **8**, 2183–2192 (2009).
135. Adams, J. & Kauffman, M. Development of the proteasome inhibitor Velcade (bortezomib). *Cancer Invest.* **22**, 304–311 (2004).
136. Gasparian, A.V. *et al.* Targeting transcription factor NF κ B: comparative analysis of proteasome and IKK inhibitors. *Cell Cycle* **8**, 1559–1566 (2009).
137. Hertlein, E. *et al.* 17-DMAG targets the nuclear factor- κ B family of proteins to induce apoptosis in chronic lymphocytic leukemia: clinical implications of HSP90 inhibition. *Blood* **116**, 45–53 (2010).
138. Wright, J.J. Combination therapy of bortezomib with novel targeted agents: an emerging treatment strategy. *Clin. Cancer Res.* **16**, 4094–4104 (2010).
139. Milhollen, M.A. *et al.* MLN4924, a NEDD8-activating enzyme inhibitor, is active in diffuse large B-cell lymphoma models: rationale for treatment of NF- κ B-dependent lymphoma. *Blood* **116**, 1515–1523 (2010).
140. Kanarek, N., London, N., Schueler-Furman, O. & Ben-Neriah, Y. Ubiquitination and degradation of the inhibitors of NF- κ B. *Cold Spring Harb. Perspect. Biol.* **2**, a000166 (2010).
141. Kanarek, N. *et al.* Spermatogenesis rescue in a mouse deficient for the ubiquitin ligase SCF β -TrCP by single substrate depletion. *Genes Dev.* **24**, 470–477 (2010).
142. Lavon, I. *et al.* Nuclear factor- κ B protects the liver against genotoxic stress and functions independently of p53. *Cancer Res.* **63**, 25–30 (2003).
143. Rothwell, P.M. *et al.* Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. *Lancet* **377**, 31–41 (2011).
144. Midgley, R.S. *et al.* Phase III randomized trial assessing rofecoxib in the adjuvant setting of colorectal cancer: final results of the VICTOR trial. *J. Clin. Oncol.* **28**, 4575–4580 (2010).
145. Coussens, L.M. & Werb, Z. Inflammation and cancer. *Nature* **420**, 860–867 (2002).
146. Karin, M. & Greten, F.R. NF- κ B: linking inflammation and immunity to cancer development and progression. *Nat. Rev. Immunol.* **5**, 749–759 (2005).
147. Hanahan, D. & Weinberg, R.A. Hallmarks of cancer: the next generation. *Cell* **144**, 646–674 (2011).
148. Berenblum, I. Challenging problems in cocarcinogenesis. *Cancer Res.* **45**, 1917–1921 (1985).
149. Endo, Y., Marusawa, H. & Chiba, T. Involvement of activation-induced cytidine deaminase in the development of colitis-associated colorectal cancers. *J. Gastroenterol.* **46** (Suppl 1), 6–10 (2011).
150. Elyada, E. *et al.* CK1 α ablation highlights a critical role for p53 in invasiveness control. *Nature* **470**, 409–413 (2011).
151. Yin, J. *et al.* p53 point mutations in dysplastic and cancerous ulcerative colitis lesions. *Gastroenterology* **104**, 1633–1639 (1993).

Copyright of Nature Immunology is the property of Nature Publishing Group and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.